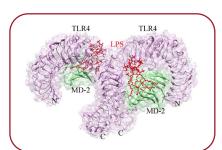


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LPS (Universal) from S. abortus equi (S-form) TLRpure[™] Sterile Solution

Source	Lizzzzhanada (LDC) fram Calmonalla abartua agui Cituzzi (amaath (uild toor)) LDC
	Lipopolysaccharide (LPS) from Salmonella abortus equi, S-type (smooth/wild-type) LPS
Concentration	Img/ml stabilised in sterile, double-distilled water (ddWater), without any additives
TLRpure™	No detectable TLR4 <i>independent</i> activity as determined by a mouse macrophage cell culture cytokine secretion assay using TLR4 deficient versus wild-type cells: standardised potent TLR4-specific agonist
Purity	Ultrapure. No detectable DNA, RNA and protein traces.
Purification Method	S-type LPS was isolated by a modified phenol-chloroform-petroleum ether method. Semi-purified LPS was subjected to further re-extraction cycles and ultracentrifugation steps, extensively electrodialysed to yield TLRpure [™] LPS.
	Filter method: certified according to Ph. Eur. 9.
Sterility	Passed according to specification:
	• No growth in Thioglycolate medium at 30-35°C after 14 days.
Endotoxin Content	 No growth in Soybean Casein Digest Broth (TSB) at 20-25°C after 14 days. Bacterial Endotoxin Test (kinetic turbidimetric LAL method) certified according to Ph. Eur. 9.
	Endotoxin Content: >5,000,000 [EU/ml].
Appearance Handling	Colourless, clear, aqueous solution Keep sterile. Prepare aliquots or working dilutions from pre-warmed (~40°C) LPS stock solution just prior to use. Ready-made solution is cell culture-grade. To yield a 100µg/ml (100x) stock solution, add 100µl of LPS to 900µl endotoxin-free and sterile ddWater (Cat. No.: IAX-900-002), or 0.9% NaCl Solution (Cat. No.: IAX-900-003) or PBS (Cat. No.: IAX-900-001) and mix well.
Activity	Optimal concentration is dependent upon cell type, species, desired activation and analysis: 0.01-1.0µg/ml <i>in vitro</i> and 5-15mg/kg <i>in vivo</i> in animal rodent models. Does not activate any TLR other than TLR4 as tested up to 10µg/ml in relevant cellular systems (mouse macrophages).
Shipping	Ambient
Storage	2-8°C
Stability	2 years after receipt (unopened and as supplied). Diluted solutions are stable for 12 hours at 2-8°C.
	Available on request

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Cat. No.: IAX-100-009	Lot. No.:
S. abortus equi TLRpure™ LPS i	s a TLR4 specific agonist
Mouse WT Macrophage 8000 7000 TLR4 KO 5000 1000 Salmonella Abortus equi LPS Mouse WT Macrophage	FIGURE: Macrophages from wild-type (WT) TLR4 expressing or TLR4 deficient (TLR4 KO) mice were stimulated with 1µg/ml TLRpure [™] S. abortus equi S-LPS. Cell culture supernatants were analysed by ELISA for IL-6 after 24h. Optimal concentrations required for activation depend upon cell species (murine, human, others), cell culture conditions (FCS concentration), sampling time and cytokine analysis. Recommended range for S-type (wild-type) LPS: 0.01-1.0µg/ml.
• Product Description	TLRpure [™] LPS has been purified according to an optimized and proprietary extraction and purification protocol, but based upon the methods published by Galanos et al. (laboratory of Westphal and Lüderitz, Freiburg, Germany). TLRpure [™] LPS lacks any detectable bacterial, (lipo-)protein, RNA or DNA or other TLR-stimulating activity due to its ultra-purified formulation. Its unique potency and purity are quality controlled using a physiological system of primary innate immune cells and a relevant biological cytokine expression read-out. Due to its amphipatic structure and strong tendency to form micelles, the generation of LPS, which is devoid of any non-TLR4 dependent immune modulatory activity, presents a major biochemical purification and analytical challenge. All immunological activity of TLRpure [™] LPS is exclusively dependent upon the presence of TLR4 as determined by the use of the corresponding control cells, derived from TLR4 deficient (TLR4 knock-out, KO) mice. TLRpure [™] LPS convenient ready-made stabilised solution makes it the reagent of choice for <i>in vitro</i> and <i>in vivo</i> experiments for superior reproducible and comparable results. These unique LPS preparations have been used in numerous publications since 1969. Compared to conventional (semipurified) LPS preparations, this low yield TLRpure [™] LPS is produced on an industrial fermentation scale under precisely controlled growth conditions to yield large batch sizes, thus allowing custom formulations/packaging.
Product Specific References	Attachment to erythrocytes of uniform salt forms of lipopolysaccharides from Salmonella abortus-equi and
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General Information	The innate immune response to lipopolysaccharide is essential for host defense against Gram-negative bacteria. In response to bacterial infection, the TLR4/MD-2 complex that is expressed on the surface of macrophages, monocytes, dendritic, and epithelial cells senses picomolar concentrations of endotoxic LPS and triggers the production of various pro-inflammatory mediators. Activation of cells by LPS is mediated on the plasma membrane by the Toll-like receptor 4 (TLR4), a member of the highly conserved protein family of TLRs, which are specialised in the recognition of microbial components. In mice, defects in TLR4 result in LPS unresponsiveness. For optimal interaction with LPS, TLR4 requires association with myeloid differentiation protein 2 (MD-2). According to current consensus activation of TLR4 is preceded by the transfer of LPS to membrane-bound (m) or soluble (s) CD14 by LPS-binding protein (LBP). This mechanism is believed to be generally true for LPS signaling. Re-form LPS and lipid A, but not S-form LPS, are capable of inducing TNF-a responses also in the absence of CD14. LPS, synthesized by most wild-type (WT) Gram-negative bacteria (S-form LPS), consists of three regions, the O-polysaccharide chain, which is made up of repeating oligosaccharide units, the core oligosaccharide chain. Furthermore, the core-oligosaccharide may be present in different degrees of completion, depending on the class (Ra to Re) to which the mutant belongs. Notably, LPS from WT bacteria are always highly heterogeneous mixtures of S-form LPS and using usyning proportion of R-form molecules lacking the O-specific chain. LPS are amphipathic molecules whose hydrophobicity decreases with increasing length of the sugar part. Based upon these differences, S- and R-form LPS show marked differences in the kinetics of their blood clearance and cellular uptake as well as in the ability to induce oxidative burst in human granulocytes and to activate the host complement system. In addition, LPS from extracellular bacteria which is eithe

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