



PRODUCT DATA SHEET

Page 1/3

Lipid A from S. minnesota R595 (Re) TLRpure[™] Sterile Solution

Cat. No.: IAX-100-00	DI Lot. No.:
Source	Lipid A derived from S. minnesota R595 (Re) TLRpure [™] LPS
Concentration	lmg/ml (0.5mg/ml for 250µg size) stabilised in sterile, double-distilled water (ddWater) without any additives
TLRpure™	No detectable TLR4 independent activity: standardised potent TLR4-specific agonist
Purity	Ultrapure. No detectable DNA, RNA and protein traces.
Purification Method	R-type (mutant/rough) LPS was isolated by a phenol-chloroform-petroleum-ether method. Semi-purified LPS was subjected to further re-extraction cycles and ultracentrifugation steps, extensively electrodialysed to yield TLRpure [™] LPS, from which Lipid A was generated by mild acid hydrolysis.
Sterility	Filter method: according to Ph. Eur. 9. Passed according to specification: • No growth in Thioglycolate medium at 30-35°C after 14 days. • No growth in Soybean Casein Digest Broth (TSB) at 20-25°C after 14 days.
Endotoxin Content	Bacterial Endotoxin Test (kinetic turbidimetric LAL method) according to Ph. Eur. 9. Endotoxin Content: >5,000,000 [EU/ml].
Appearance	Colourless, clear, aqueous solution
Handling	Keep sterile. Prepare working dilutions from pre-warmed (~40°C) Lipid A stock solution just prior to use. Ready-to-use, sterile stock solution is cell culture-grade. No solubilisation required. Do not pre-dilute in buffer (e.g. PBS) as this will lead to precipitation of Lipid A. To yield a 100µg/ml (1,000-100x) stock solution add 100µl of Lipid A to 900µl endotoxin-free and sterile ddWater (Cat. No.: IAX-900-002) (not PBS) and mix well.
Activity	Optimal concentration is dependent upon cell type, species, desired activation and analysis: 0.1-1.0µg/ml <i>in vitro</i> and 5-15mg/kg <i>in vivo</i> in animal rodent models. Does not activate any TLR other than TLR4 as tested up to 1µg/ml in relevant cellular systems (mouse macrophages).
Shipping	Ambient
Storage	2-8°C
Stability	 Upon receipt, store product at 2-8°C. Do not freeze. In its unopened original vial, the product is stable for at least 24 months when stored at 2-8°C. Once the glass vial is opened, or if aliquoted into sterile vials under sterile conditions, the product remains stable for an additional 12 months at 2-8°C. Pre-diluted, sterile aqueous solutions (e.g., 10-100µg/ml) are stable for a maximum of 12 hours when stored at 2-8°C due to vial surface effects on diluted solutions.
MSDS	Available on request

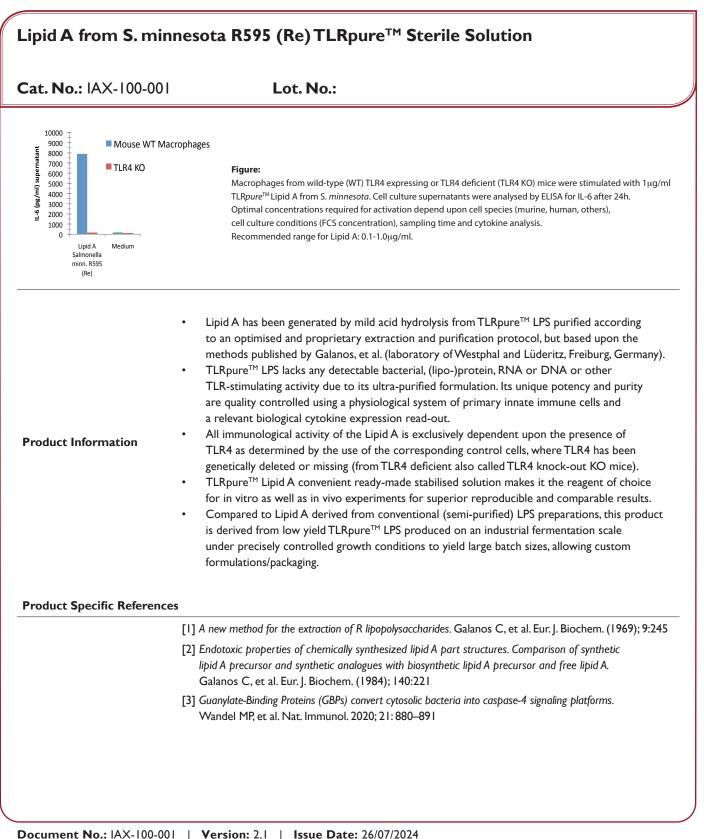
Document No.: IAX-100-001 | Version: 2.1 | Issue Date: 26/07/2024

DISCLAIMER: THIS PRODUCT IS NOT INTENDED OR APPROVED FOR HUMAN, DIAGNOSTICS OR VETERINARY USE. USE OF THIS PRODUCT FOR HUMAN OR ANIMAL TESTING MAY BE EXTREMELY HAZARDOUS AND MAY RESULT IN DISEASE, SEVERE INJURY, OR DEATH. THIS PRODUCT IS FOR RESEARCH USE ONLY (RUO). MATERIAL SAFETY DATA: This material should be considered hazardous until information to the contrary becomes available. Do not ingest, swallow, inhale or get into the blood stream. Do not get in eyes, on skin, or clothing. Wash thoroughly after handling. This information contains some, but not all, of the information required for the safe and proper use of this material. Access to this material must be restricted to personnel, who is appropriately experienced, qualified, competent and properly trained to use it. Material Safety Data Sheet is available upon request.





PRODUCT DATA SHEET) Page 2/3



DISCLAIMER: THIS PRODUCT IS NOT INTENDED OR APPROVED FOR HUMAN, DIAGNOSTICS OR VETERINARY USE. USE OF THIS PRODUCT FOR HUMAN OR ANIMAL TESTING MAY BE EXTREMELY HAZARDOUS AND MAY RESULT IN DISEASE, SEVERE INJURY, OR DEATH. THIS PRODUCT IS FOR RESEARCH USE ONLY (RUO). MATERIAL SAFETY DATA: This material should be considered hazardous until information to the contrary becomes available. Do not ingest, swallow, inhale or get into the blood stream. Do not get in eyes, on skin, or clothing. Wash thoroughly after handling. This information contains some, but not all, of the information required for the safe and proper use of this material. Access to this material must be restricted to personnel, who is appropriately experienced, qualified, competent and properly trained to use it. Material Safety Data Sheet is available upon request.



Innaxon United Kingdom Email: info@innaxon.com Website: www.innaxon.co.uk



PRODUCT DATA SHEET

Page 3/3

 Activation of cells by LPS is mediated by the Toll-like receptor 4 (TLR4), a member of the highly conserved protein family of TLRs, which are specialised in the recognition of microbial components. In mice, defects in TLR4 result in LPS unresponsiveness. For optimal interaction with LPS, TLR4 requires association with mysloid differentiation protein 2 (MD-2). According to current consensus activation of TLR4 is preceded by the transfer of LPS to membrane-bound (m) or soluble (s) CD14 by LPS-binding protein (LBP). This mechanism is believed to be generally true for LPS signaling. Re-form LPS, consists of three regions, the O-polysaccharide chain, which its made up of repeating oligosaccharide units, the core oligosaccharide and the lipid A, which harbors the endotoxic activity of the entire molecule. R-form LPS synthesised by the so-called rough (R) mutants of Gram-negative bacteria lacks the O-specific chain. Furthermore, the core-oligosaccharide may be present in different degrees of completion, depending on the class (Ra to Re) to which the mutant belongs. Monophosphoryl Lipid A (MPLA) represents a detoxified derivative of Lipid A and constitutes an important adjuvant in prophylactic and therapeutic vaccines. References [1] <i>R-form LPS, the master key to the activation of TLR4/MD-2-positive cells</i>. Huber M, et al. Eur. J. Immunol. (2006); 36-701 [2] CD14 is required for MybB8-independent LPS signaling. Jiang Z, Georgel P, Du X, Shamel L, Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Bauter B, Nat. Immunol. (2005); 56-56 [3] <i>Defective LPS signaling in C3HIHelg and C3TBL/10ScCr mice: mutations in Tir4 gene.</i> Poltorak A, He X, Smirnova I, Liu MY, Van Huiffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Cassagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] <i>Structural relationship of Salmonella 0 and R antgens.</i> Lideritz O, Galanos C, et al. Ann. N.Y. Acad. S	Cat. No.: IAX-100-001	Lot. No.:
 [1] R-form LPS, the master key to the activation of TLR4/MD-2-positive cells. Huber M, et al. Eur. J. Immunol. (2006); 36:701 [2] CD14 is required for MyD88-independent LPS signaling. Jiang Z, Georgel P, Du X, Shamel L, Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Beutler B. Nat. Immunol. (2005); 6:565 [3] Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] Preparation and properties of antisera against the lipid-A component of bacterial lipopolysaccharides. Galanos C, et al. Lur. J. Biochem. (1971); 24:116 [6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of 	General Information	 highly conserved protein family of TLRs, which are specialised in the recognition of microbial components. In mice, defects in TLR4 result in LPS unresponsiveness. For optimal interaction with LPS, TLR4 requires association with myeloid differentiation protein 2 (MD-2). According to current consensus activation of TLR4 is preceded by the transfer of LPS to membrane-bound (m) or soluble (s) CD14 by LPS-binding protein (LBP). This mechanism is believed to be generally true for LPS signaling. Re-form LPS and lipid A, but not S-form LPS, are capable of inducing TNF-α responses also in the absence of CD14. LPS, synthesised by most wild-type (WT) Gram-negative bacteria (S-form LPS), consists of three regions, the O-polysaccharide chain, which is made up of repeating oligosaccharide units, the core oligosaccharide and the lipid A, which harbors the endotoxic activity of the entire molecule. R-form LPS synthesised by the so-called rough (R) mutants of Gram-negative bacteria lacks the O-specific chain. Furthermore, the core-oligosaccharide may be present in different degrees of completion, depending on the class (Ra to Re) to which the mutant belongs. Monophosphoryl Lipid A (MPLA) represents a detoxified derivative of Lipid A and constitutes
 Eur. J. Immunol. (2006); 36:701 [2] <i>CD14 is required for MyD88-independent LPS signaling</i>. Jiang Z, Georgel P, Du X, Shamel L, Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Beutler B. Nat. Immunol. (2005); 6:565 [3] <i>Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene</i>. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] <i>Structural relationship of Salmonella 0 and R antigens</i>. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] <i>Preparation and properties of antisera against the lipid-A component of bacterial lipopolysaccharides</i>. Galanos C, et al. Eur. J. Biochem. (1971); 24:116 [6] <i>Lipid A: chemical structure and biological activity</i>. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] <i>Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of</i> 	References	[1] R-form LPS. the master key to the activation of TLR4/MD-2-bositive cells. Huber M. et al.
 Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Beutler B. Nat. Immunol. (2005); 6:565 [3] Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] Preparation and properties of antisera against the lipid-A component of bacterial lipopolysaccharides. Galanos C, et al. Eur. J. Biochem. (1971); 24:116 [6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of 		
 He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] Preparation and properties of antisera against the lipid-A component of bacterial lipopolysaccharides. Galanos C, et al. Eur. J. Biochem. (1971); 24:116 [6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of 		Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Beutler B.
 [4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] Preparation and properties of antisera against the lipid-A component of bacterial lipopolysaccharides. Galanos C, et al. Eur. J. Biochem. (1971); 24:116 [6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of 		He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C,
Galanos C, et al. Eur. J. Biochem. (1971); 24:116 [6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of		[4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y.
[7] Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of		
		[6] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17

Document No.: IAX-100-001 | Version: 2.1 | Issue Date: 26/07/2024

DISCLAIMER: THIS PRODUCT IS NOT INTENDED OR APPROVED FOR HUMAN, DIAGNOSTICS OR VETERINARY USE. USE OF THIS PRODUCT FOR HUMAN OR ANIMAL TESTING MAY BE EXTREMELY HAZARDOUS AND MAY RESULT IN DISEASE, SEVERE INJURY, OR DEATH. THIS PRODUCT IS FOR RESEARCH USE ONLY (RUO). MATERIAL SAFETY DATA: This material should be considered hazardous until information to the contrary becomes available. Do not ingest, swallow, inhale or get into the blood stream. Do not get in eyes, on skin, or clothing. Wash thoroughly after handling. This information contains some, but not all, of the information required for the safe and proper use of this material. Access to this material must be restricted to personnel, who is appropriately experienced, qualified, competent and properly trained to use it. Material Safety Data Sheet is available upon request.