





PRODUCT DATA SHEET ) Page 1/3

## LPS from S. minnesota R60 (Ra) TLRpure<sup>™</sup> Sterile Solution

Cat. No.: IAX-100-0	16 Lot. No.:
Source	Lipopolysaccharide (LPS) from S. minnesota strain R60 (Ra), R-type (rough/mutant) LPS
Concentration	Img/ml stabilised in sterile, double-distilled water (ddWater), without any additives
TLRpure™	No detectable TLR4 <i>independent</i> activity as determined by a mouse macrophage cell culture cytokine secretion assay using TLR4 deficient versus wild-type cells: standardised potent TLR4-specific agonist
Purity	Ultrapure. No detectable DNA, RNA and protein traces.
Purification Method	R-type (mutant/rough) LPS was isolated by phenol-chloroform-petroleum-ether method. Semi-purified LPS was subjected to further re-extraction cycles and ultracentrifugation steps, extensively electrodialysed to yield TLRpure <sup>™</sup> LPS.
Sterility	Filter method: certified according to Ph. Eur. 9. Passed according to specification: • No growth in Thioglycolate medium at 30-35°C after 14 days. • No growth in Soybean Casein Digest Broth (TSB) at 20-25°C after 14 days.
Endotoxin Content	Bacterial Endotoxin Test (kinetic turbidimetric LAL method) certified according to Ph. Eur. 9. Endotoxin Content: >10,000,000 [EU/ml].
Appearance	Colourless, clear, aqueous solution
Handling	<ul> <li>Keep sterile. Prepare working dilutions from pre-warmed (~40°C) LPS stock solution just prior to use. Ready-to-use, sterile stock solution is cell culture-grade. No solubilisation required.</li> <li>To yield a 100µg/ml (1,000-100x) stock solution add 100µl of LPS to 900µl endotoxin-free and sterile ddWater (Cat. No.: IAX-900-002) or 0.9% NaCl Solution (Cat. No.: IAX-900-003) and mix well.</li> </ul>
Activity	Optimal concentration is dependent upon cell type, species, desired activation and analysis: 0.01-1.0µg/ml. Does not activate any TLR other than TLR4 as tested up to 1µg/ml in relevant cellular systems (mouse macrophages).
Shipping	Ambient
Storage	2-8°C
Stability	<ul> <li>Upon receipt, store product at 2-8°C.</li> <li>Do not freeze.</li> <li>In its unopened original vial, the product is stable for at least 24 months when stored at 2-8°C.</li> <li>Once the glass vial is opened, or if aliquoted into sterile vials under sterile conditions, the product remains stable for an additional 12 months at 2-8°C.</li> <li>Pre-diluted, sterile aqueous solutions (e.g., 10-100µg/ml) are stable for a maximum of 12 hours when stored at 2-8°C due to vial surface effects on diluted solutions.</li> </ul>
MSDS	Available on request

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## PRODUCT DATA SHEET

**Page** 2/3

<ul> <li>1LR4 as determined by the use of the corresponding control cells, where 1LR4 has been genetically deleted or missing (from TLR4 deficient also called TLR4 knock-out KO mice).</li> <li>TLRpure<sup>™</sup> LPS convenient ready-made stabilised solution makes it the reagent of choice for in vitro as well as in vivo experiments for superior reproducible and comparable results.</li> <li>Compared to LPS derived from conventional (semi-purified) LPS preparations, this product is derived from low yield TLRpure<sup>™</sup> LPS produced on an industrial fermentation scale under precisely controlled growth conditions to yield large batch sizes, allowing custom formulations/packaging.</li> </ul>	at. No.: IAX-100-016	b Lot. No.:
<ul> <li>[2] Electrodialysis of lipopolysaccharides and their conversion to uniform salt forms. Galanos C &amp; Lüderitz O. Eur. J. Biochem. (1975); 54:603</li> <li>[3] Induction of human granulocyte chemiluminescence by bacterial lipopolysaccharides. Kapp A, Freudenberg M, Galanos C. Infect. Immun. (1987); 55:758</li> <li>[4] Isolation and purification of R-form lipopolysaccharides. Galanos C &amp; Lüderitz O. Methods in Carbohydrate Chem. (1993); 9:11</li> <li>[5] Guanylate-Binding Proteins (GBPs) convert cytosolic bacteria into caspase-4 signaling platforms. Wandel MP, et al. Nat. Immunol. 2020; 21: 880–891</li> <li>[6] Synthetic glycolipid-based TLR4 antagonists negatively regulate TRIF-dependent TLR4 signalling in human macrophages. Palmer C, Facchini F A, Jones R PO, Neumann F, Peri F, Pirianov G.</li> </ul>	Product Information	<ul> <li>protocol, but based upon the methods published by Galanos, et al. (laboratory of Westphal and Lüderitz, Freiburg, Germany).</li> <li>TLRpure<sup>™</sup> LPS lacks any detectable bacterial, (lipo-)protein, RNA or DNA or other TLR-stimulating activity due to its ultra-purified formulation. Its unique potency and purity are quality controlled using a physiological system of primary innate immune cells and a relevant biological cytokine expression read-out.</li> <li>All immunological activity of the Lipid A is exclusively dependent upon the presence of TLR4 as determined by the use of the corresponding control cells, where TLR4 has been genetically deleted or missing (from TLR4 deficient also called TLR4 knock-out KO mice).</li> <li>TLRpure<sup>™</sup> LPS convenient ready-made stabilised solution makes it the reagent of choice for in vitro as well as in vivo experiments for superior reproducible and comparable results.</li> <li>Compared to LPS derived from conventional (semi-purified) LPS preparations, this product is derived from low yield TLRpure<sup>™</sup> LPS produced on an industrial fermentation scale under precisely controlled growth conditions to yield large batch sizes, allowing custom</li> </ul>
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Page 3/3



## **PRODUCT DATA SHEET** )

LPS from S. minnesota R60 (Ra) TLRpure<sup>™</sup> Sterile Solution Cat. No.: IAX-100-016 Lot. No.: Activation of cells by LPS is mediated by the Toll-like receptor 4 (TLR4), a member of the highly conserved protein family of TLRs, which are specialised in the recognition of microbial components. In mice, defects in TLR4 result in LPS unresponsiveness. For optimal interaction with LPS, TLR4 requires association with myeloid differentiation protein 2 (MD-2). According to current consensus activation of TLR4 is preceded by the transfer of LPS to membrane-bound (m) or soluble (s) CD14 by LPS-binding protein (LBP). This mechanism is believed to be generally true for LPS signaling. Re-form LPS and lipid A, but not S-form LPS, are capable of inducing TNF-a responses also in the absence of CD14. LPS, synthesised by most wild-type (WT) Gram-negative bacteria (S-form LPS), consists of three regions, the O-polysaccharide chain, which is made up of repeating oligosaccharide **General Information** units, the core oligosaccharide and the lipid A, which harbors the endotoxic activity of the entire molecule. R-form LPS synthesised by the so-called rough (R) mutants of Gram-negative bacteria lacks the O-specific chain. Furthermore, the core-oligosaccharide may be present in different degrees of completion, depending on the class (Ra to Re) to which the mutant belongs. Notably, LPS from WT bacteria are always highly heterogeneous mixtures of S-form LPS molecules containing I to over 50 repeating oligosaccharide units and contain ubiquitously a varying proportion of R-form molecules lacking the O-specific chain. LPS are amphipathic molecules whose hydrophobicity decreases with increasing length of the sugar part. Based upon these differences, S- and R-form LPS show marked differences in the kinetics of their blood clearance and cellular uptake as well as in the ability to induce oxidative burst in human granulocytes and to activate the host complement system. References [1] R-form LPS, the master key to the activation of TLR4/MD-2-positive cells. Huber M, et al. Eur. J. Immunol. (2006); 36:701 [2] CD14 is required for MyD88-independent LPS signaling. Jiang Z, Georgel P, Du X, Shamel L, Sovath S, Mudd S, Huber M, Kalis C, Keck S, Galanos C, Freudenberg M, Beutler B. Nat. Immunol. (2005); 6:565 [3] Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, Freudenberg M, Ricciardi-Castagnoli P, Layton B, Beutler B. Science (1998); 282:2085 [4] Structural relationship of Salmonella 0 and R antigens. Lüderitz O, Galanos C, et al. Ann. N.Y. Acad. Sci. (1966); 133:349 [5] Lipid A: chemical structure and biological activity. Lüderitz O, Galanos C, et al. J. Infect. Dis. (1973); 128:17 [6] Defective immunogenic cell death of HMGB1-deficient tumors: compensatory therapy with TLR4 agonists. Yamazaki T, et al. Cell Death and Differentiation (2014); 21:69

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